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Courses » Applications of interactomics using Genomics and proteomics technologies

Announcements Course Ask a Question Progress FAQ

Unit 4 - Week 3

Register for Certification exam

Course outline

How to access the portal

Week 1

Week 2

Week 3

- Lecture 11 :
 Using
 functional
 proteomics to
 identify
 biomarkers and
 therapeutic
 targets-II
- Lecture 12:
 Applications of protein microarrays in Malaria
 Research-I
- Lecture 13: Applications of protein microarrays in Malaria Research-II
- Applications of protein microarrays in Cancer Research-I

Assignment 3

The due date for submitting this assignment has passed.

As per our records you have not submitted this assignment.

Due on 2019-03-20, 23:59 IST.

- 1) When you scan your microarray slide, you see several saturated spots. What will you do in **1 point** such a case?
 - Increase PMT settings so that more spots become saturated
 - Lower PMT settings to reduce spot intensity
 - Increase PMT settings to increase the number of bright spots
 - Lower PMT settings to lower the background signals

No, the answer is incorrect.

Score: 0

Accepted Answers:

Lower PMT settings to reduce spot intensity

- 2) Which of the following options are in the correct order in case of a microarray experiment. 1 point
 - Normalization----Power calculation----Image processing---- PCA plot
 - Power calculation----Image processing---Normalization---PCA plot
 - Image processing---- PCA plot---Normalization---Power calculation
 - PCA plot---Image processing---Power calculation---Normalization

No, the answer is incorrect.

Score: 0

Accepted Answers:

Power calculation----Image processing---Normalization----PCA plot

3) Imagine that you are performing a microarray experiment where chips are probed with **1 point** dengue positive patient sera to study IgG responses? You plan to use the slide shown below which can probe 4 patient sera simultaneously. Viral proteins are printed on the slide as *In vitro* transcription and translation (IVTT) spots just like the Nucleic Acid Programmable Arrays (NAPPA). The slide also has several other control spots. Answer the following questions (Q3 to Q6) based on your experiment.

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Assignment 3	
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Assignment 3: Solutions	
Week 4	
Week 5	
Week 6	3) Based on the printed spots on your slide, which one of the following can NOT be a positive control in your experiment?
Week 7	
Week 8	IgG spots Human IgG spots
	Purified viral protein spots
Interaction Session	IVTT spots
	No, the answer is incorrect.
	Score: 0
	Accepted Answers: IVTT spots
	4) What else can you study using a similar microarray slide and patient samples? 1 point
	Only IgG responses in patients
	IgG responses and protein interactions
	Depends on the antibody
	Depends on the detection system
	No, the answer is incorrect. Score: 0
	Accepted Answers:
	Depends on the antibody
	5) What will you do after you finish your experiment and the slides are dry? 1 point
	You will use a GPR file to scan your slides
	You will use the Gal file that you created to scan your slides
	You will export the GPR file provided by the manufacturer as an excel sheet
	You will use a Gal file after scanning your slides
	No, the answer is incorrect. Score: 0
	Accepted Answers: You will use a Gal file after scanning your slides
	6) While you are performing the experiment, you forget one main step. After scanning, you 1 point see that your slide has very faint signals for all 4 patients. Which step could that most likely be from the options below?
	You forgot to add secondary antibody
	You did not wash your slides after addition of tertiary antibody
	You did not incubate the sera with E. coli lysate before hybridization
	You did not cover your slides after addition of tertiary antibody

No, the answer is incorrect. Score: 0 **Accepted Answers:** You did not cover your slides after addition of tertiary antibody 7) While analyzing your data you realize that you need to include an important patient data which you did not add earlier. This data will help you segregate the sample population into two groups. What will you do? You will rescan the slides You will classify the samples into two groups and redo the hybridization You will map the new patient data to the existing datasheet and re-analyze it You cannot include this data in your study anymore No, the answer is incorrect. Score: 0 **Accepted Answers:** You will map the new patient data to the existing datasheet and re-analyze it 8) Answer Question 8 and 9 are based on the information given below: 1 point Phosphorylation of PLCy1 at Tyr8783 activates the enzymatic activity of PLCy1 which is crucial event of signal transduction of T-lymphocytes upon stimulation of T-cell receptors. In order to map the downstream signalling pathways of activated PLCy1, Chan et al., stimulated the Jurkat T lymphocytes and J.gamma1 cells, a mutant line of Jurkat T cells deficient in PLCy1 with antibodies to CD3 + CD28 and compared the phosphorylation kinetics of four signalling proteins (PLCy1, Akt, p44/42 MAPK and MEK1/2). The phosphorylation level of all the four proteins over the period of 30 min is provided in the graph below. Analyse the graph given below and answer question 8 and 9. phospho-PLC_γ (Tyr 783) phospho-Akt (Ser 473) 1.0 1.5 0.8 Fold change (log₂) 1.2 0.6 0.9 0.4 0.6 -J. gamma1 0.2 0.3 Wild-type 0.0 0.0 15 25 10 15 25 20 phospho-p44/42 MAPK phospho-MEK1/2 3.5 2.0 3.0 Fold change (log₂) 1.6 2.5-1.2 2.0 1.5 1.0 0.4 0.5 0.0 15 25 ò Time (min) Time (min) 8) Despite knowing the fact that the J.gamma1 cell lines are deficient in PLCy1, phosphorylation kinetics of PLCy1 was studied in both the cell lines and probed the lysates with phospho- PLCy1. What could be the plausible reason? To check if the phospho-PLCy1 antibody is cross-reacting with other proteins To ensure that J.gamma1 cell lines are deficient in PLCy1 To check the levels of PLCy1 in both the cell lines

To study the downstream signalling kinetics



