LECTURE-36

Salivary Proteomics

<u>Handout</u>

PREAMBLE

Proteins are the effector molecules of cellular pathways and its over-expression or deficiency results in diseases. There are many diseases which are caused due to aberrant protein-protein interactions, which are indispensible for normal functioning of cells. Thus, proteomic profiling is very common approach to find out modulation of proteins due to any disease. For comparative proteomic analysis, samples are taken from the normal and diseased individuals. These samples typically include serum or plasma, tissue and cerebro-spinal fluid. Obtaining these samples need invasive procedures – like a lumbar puncture or sometimes surgery. Although these are useful sample but amount obtained is very less and involves invasive procedure. Therefore, newer samples need to be discovered for protein profiling, which can be obtained through non-invasive techniques. One such sample is saliva, which is abundant, non-invasive and very easily obtained without any harm to the patient. Thus, if a biomarker could be discovered in a bodily fluid like saliva or urine, it will be much easier to obtain samples to identify the occurrence and progression of disease in an individual.

OUTLINE

- I. Introduction
- II. Collection and processing of saliva samples for diagnostic and research purposes
- III. Methodology for protein extraction from saliva
- IV. Applications of salivary proteomics
- V. Advantages and challenges
- VI. Conclusions

I. INTRODUCTION

Saliva is a colorless fluid secreted by the salivary glands of the oral cavity. There are three main salivary glands in humans: the parotid glands, the sub-mandibular glands and sub-lingual glands, along with numerous small glands. Saliva is a fluid which helps to start the preliminary digestion of food as it has enzymes such as salivary amylase. The saliva composition is mainly as follows: 99.5% water, electrolytes, mucus, enzymes, anti-bacterial agents. The electrolytes include sodium, potassium, magnesium, bicarbonates, chlorides and phosphates etc. (Fig 1).



Fig 1. Composition of saliva

Many proteins have been detected in saliva and most abundant protein is salivary amylase. Other proteins which have been discovered in saliva include; Histatins, Statherins, Lysozyme, Amylases, Perioxidases, Mucins, Lactoferrin, Kallikarein, Defensins, Cystatins, Basic PRP, Peroxidase, S-IgA and Agglutinin etc. (Fig 2).

NPTEL WEB COURSE – ADVANCED CLINICAL PROTEOMICS



Fig 2. Six major types of salivary proteins: histatins, proline-rich proteins (acidic, basic and glycosylated), statherins and cystatins.

These salivary proteins have various functions such as digestion of food, antimicrobial activity, lubrication and protection of oral surfaces. Till date around 3000 proteins have been detected in saliva.

NPTEL WEB COURSE – ADVANCED CLINICAL PROTEOMICS



Fig. 3. Major proteins found in human saliva (E. Scarano et al. 2010)

II. COLLECTION OF SALIVA SAMPLES FOR DIAGNOSTIC AND RESEARCH PURPOSES

Normally in adults, the amount of saliva produced is 1 - 1.5 L. Generally, this is produced at the rate of 1 - 3 mL per minute, when production of salivary fluid is stimulated. Saliva is mainly contributed by three salivary glands; the gingival fold, oral mucosa transudate, mucous layer of the nasal cavity and pharynx.

Standard protocol for collection of saliva for diagnostic purposes

- Saliva collection is generally performed in the morning, optimally around 10-11 AM
- The subject is not allowed to eat, drink or conduct oral hygiene procedures such as brushing teeth at least 1 hour prior to the sample collection.
- 3. The subject is given drinking water for a mouth rinse.
- After five minutes of the mouth rinse, the subject should be asked to spit whole saliva (WS) into a 50 ml sterile Falcon tube.
- 5. Talking should be ideally avoided before and during sample collection.
- The subject should spit into the collection tube about once a minute for upto 10 minutes. At times, the subject is given citric acid solution rinse, and then the saliva is collected.
- 7. Ideally, around 5 ml sample should be collected at one time.

Processing of samples after collection of saliva

1. One important guideline to be strictly followed is that all collected samples must always be kept on ice prior to processing. 2. While collecting the sample, the tube should be placed on ice.

3. The first step in sample preparation is to add proteinase inhibitors. Thus, 0.2 µL proteinase inhibitor cocktail is added per 100 µl saliva.

4. Next, Na_3OV_4 is added, and mixed gently.

5. Tubes are centrifuged at 2600xg for 15 minutes at 4°C.

6. After the spin, pellet and supernatant are stored at -80°C separately.

III. Methodology for protein extraction from saliva

The methodology for extraction of proteins from saliva is similar to any other protein extraction protocol. The samples are centrifuged to remove the presence of any impurity. The remaining sample is then subjected to TCA-Acetone precipitation.

In saliva samples, amylase is the most abundant protein. Thus, many researchers first use the depletion protocol for depletion of amylase from the saliva sample.

IV. Applications - current progress in diagnostics using saliva proteomics

The main reason for studying the proteomic profile of saliva is for the discovery of biomarkers. The protein signatures that help in distinction between healthy and diseased conditions is investigated. The proteins in saliva are precipitated and proteomic analysis using gel-based (2DE, DIGE) or gel-free mass spectrometry techniques is performed. Various examples demonstrate how salivary proteome has been investigated for different applications (Fig 4).



Fig 4. Various fields in which salivary proteomics has been utilized.

(a) Diagnosis of oral diseases

The proteins in saliva can be monitored for identifying the oral diseases. In this respect, much progress has been made for identification and prognosis of the oral diseaseperiodontitis. Periodontitis affects the bones and ligaments supporting the teeth. This has lead to the formation of the OralOme, a complete collection of proteome of the saliva. The OralOme helps detect the source of the proteins, and also find out which proteins are secreted only under diseased conditions. It is a comprehensive database to facilitate discovery of biomarkers in saliva for various diseases. Also, much progress is being made to identify dental diseases, like dental carries using proteomic markers in saliva.

(b) Saliva for diagnosing cancers

Saliva has been used to diagnose cancers such as oral cancers and head and neck cancer. For breast cancer, the biomarkers screened from saliva are Vasclular Endothelial Growth Factor (VEGF), human epidermal growth factor receptor 2, epidermal growth factor (EGF) and carcinoembryogenic antigen (CEA). The salivary c-erbB-2 levels correlated strongly with breast malignancy in women (Streckfus et al. 2005). For oral squamous cell carcinoma, the salivary proteases have been used for detection of the disease. Tumor-derived exosomes and microvesicles in head and neck cancer have been suggested as useful source for biomarker discovery (Principe S, 2013). Also, the efforts are being taken to use proteomic profiling to distinguish between the malignant and non-malignant oral lesions, which develop on OSCC. The proteomic profile of patients with oral squamous cell carcinoma (OSCC) revealed higher levels of

salivary transferrin levels that correlated with size and stage of tumor (Jou et al. 2010). The salivary transcriptome has also been analyzed for diagnosing pancreatic cancer. These studies have provided potential targets, which could be potentially screened for diagnosis of cancer.

(c) For identifying microbial infections

Certain types of microbes leave certain trails, which can be identified and infection can be tracked. Some viral or bacterial infections lead to release of specific factors from the host cells. The bacteria-derived enzymes; collagen-degrading enzymes, proteases, aminopeptidases and peptidases have been detected in proteomic profiling. Thus, either the host-derived factors in response to inflammation or the bacteria-derived enzymes can serve as biomarkers in saliva for infections. This also helps in identifying the stage of infection and thus, helps to decide the therapeutics to be administered depending on stage of infection.

(d) Salivary proteomics for identification of Sjögren's syndrome

Sjögren's syndrome (SS) is an auto-immune disease in which salivary glands and lachrymal glands are destroyed by the host's immune system. Mass spectrometric analysis has been performed on salivary samples, and definite up-regulation and downregulation of certain proteins has been reported. These proteins are potential targets for early disease detection and disease progression.

(e) Animal health and nutrition in cattle and livestock animals

Saliva biomarkers can be used to monitor the disease status of animals. The proteomic analysis can also be used to make necessary diet changes or changes in environmental conditions to maintain optimum health of the animals. This will help in keeping track of animal physiology.

(f) Salivary proteomic analysis for infectious diseases

Salivary proteomics can be used to identify even infectious diseases caused by pathogens. One area in which much progress has been made is Malaria. One of the studies showed that the inflammatory and erythrocyte-derived proteins were highly up-regulated in malaria-positive patients. A proteomic profiling of depleted saliva samples from uncomplicated *Plasmodium falciparum* malaria children was performed and three proteins, PFL0480w, PF08_0054 and PFI0875w, were identified in malaria patients (Huang et al. 2012). Thus, saliva can be clinically used for malaria biomarker detection.

V. ADVANTAGES AND CHALLENGES

| ADVANTAGES | CHALLENGES |
|--|---------------------------------------|
| Completely non-invasive, painless | Very low amount of protein is present |
| procedure, no injections or surgery is | in saliva. |
| required. | |
| Extremely cost-effective as sample | Composition of saliva varies with day |
| collection requires no complex | and night time. |
| procedure or treatment on patient. | |
| Abundant sample and there is no | Composition of saliva varies with |
| restriction on sample amount, which | physiological conditions and |
| aids in protein profiling. | biochemical changes. |
| Samples can be obtained on demand | The composition of saliva has been |
| at any time without any delay. | observed to be quite different in |
| | children and adults. |
| Patient health is not compromised in | The content of saliva changes with |
| any way. | incoming oral stimuli – like eating, |
| | drinking and even brushing teeth or |
| | flossing. |

NPTEL WEB COURSE – ADVANCED CLINICAL PROTEOMICS

| Fresh sample can be obtained | Saliva has many other components |
|-------------------------------------|--|
| whenever required, without the help | other than proteins like food, |
| of a skilled technician. | microorganisms, dead cells, which |
| | may require extra washing steps to |
| | remove from required sample. |
| | |
| Easy handling of sample. | Quantity of saliva obtained during one |
| | sample collection is low. |
| | |

VI. CONCLUSIONS

- Saliva has great potential to be used for diagnostics. It can be obtained easily, non-invasively and whenever required.
- It has been used accurately to diagnose many diseases such as dental problems, oral cancers and other infectious diseases like malaria.
- Even though saliva proteomics is a field with vast possibilities; however, there are some challenges need to overcome like low amount of proteins in saliva, presence of high-abundant proteins and variations in the composition of saliva.
- With advancement in proteomic technologies such as mass spectrometry, now comprehensive profiling of salivary proteome in various diseases will be possible.
 Targets identified from salivary proteomics will play a central role for early diagnostics.

VII. REFERENCES

- Sandra K. Al-Tarawneh, Michael B. Border, Christopher F. Dibble, and SompopBencharit Defining Salivary Biomarkers Using Mass Spectrometry-Based Proteomics: A Systematic Review, OMICS A Journal of Integrative Biology, Volume 15, Number 6, 2011.
- Walter L Sequiera, Colin Dawes. The Salivary Proteome Challenges and Perspectives, Proteomics – Clinical Applications, Volume, Issue 11-12.
- Roslinda Mohamed, Jennifer-Leigh Campbell, Justin Cooper-White, GoceDimeskiandChamindiePunyadeera. The impact of saliva collection and processing methods on CRP, IgE, and Myoglobin Immunoassays, Mohamed et al. Clinical and Translational Medicine 2012, 1:19.
- Franky D. Shah, Rasheedunnisa Begum, Bhairavi N. Vajaria, Kinjal R. Patel, Jayendra B. Patel, Shilin N. Shukla, Prabhudas S. Patel, A Review on Salivary Genomics and Proteomics Biomarkers in Oral Cancer. Ind J Clin Biochem (Oct-Dec 2011) 26(4):326–334.
- K. Jessie, O.H. Hashim, Z.H.A. Rahim 2008, Protein Precipitation Method for Salivary Proteins and rehydration buffer for two dimensional electrophoresis. Biotechnology, 7: 686-693.
- E. Scarano, A. Fiorita, P.M. Picciotti, G.C. Passali, L. Calò, T. Cabras, R. Inzitari, C. Fanali, I. Messana, M. Castagnola, G. Paludetti, Proteomics of saliva: personal experience. ACTA otorhinolaryngologicaitalica 2010; 30: 125-130.

- Nicolai J Bonne and David TW Wong, Salivary Biomarker Development using genomic, proteomic and metabolomic approaches, Genome Medicine 2012, 4: 82.
- Lamy E and Mau M, Saliva Proteomics as an emerging, non-invasive tool to study livestock, physiology, nutrition and diseases. J Proteomics, 2012 Jul 19; 75 (14): 4251-8.
- Zhang A, Sun H, Wang P, Wang X, Salivary proteomics in biomedical research, ClinChimActa, 2012 Nov 9, 415C: 261-265.
- Vitorino R, Guedes S, Manadas B, Ferreiera R, Amado F, Towards a standardized saliva proteome analysis methodology, J Proteomics, 2012 Sep 18; 75(17):5140-65.
- Huang H, Mackeen MM, Cook M, Oriero E, Locke E, Thézénas ML, Kessler BM, Nwakanma D. and Casals-Pascual C and Huang et al. Proteomic identification of host and parasite biomarkers in saliva from patients with uncomplicated *Plasmodium falciparum* malaria, Malaria Journal 2012, 11:178.
- de Almeida Pdel V, Grégio AM, Machado MA, de Lima AA, Azevedo LR.
 Saliva composition and functions: a comprehensive review. J Contemp Dent Pract. 2008 Mar 1;9(3):72-80.
- Yamada N, Yuji R, Suzuki E, The Current status and future prospects of the salivary proteome, Journal of health science, 55(5) 682-688 (2009).
- Castagnola M et al, Expert Rev Proteomics, 2012;9(1):33-46. The Human Salivary Proteome.

- Amado FM, Ferreira RP, Vitorino R. One decade of salivary proteomics: Current approaches and outstanding challenges. Clin Biochem. 2013 Apr;46(6):506-17.
- Principe S, Hui AB, Bruce J, Sinha A, Liu FF, Kislinger T. Tumor-derived exosomes and microvesicles in head and neck cancer: implications for tumor biology and biomarker discovery. Proteomics. 2013 Mar 18. doi: 10.1002/pmic.201200533.
- Jou YJ, Lin CD, Lai CH, Chen CH, Kao JY, Chen SY, Tsai MH, Huang SH, Lin CW. Proteomic identification of salivary transferrin as a biomarker for early detection of oral cancer. Anal Chim Acta. 2010 Nov 29;681(1-2):41-8.
- Streckfus C, Bigler L. The use of soluble, salivary c-erbB-2 for the detection and post-operative follow-up of breast cancer in women: the results of a five-year translational research study. Adv Dent Res. 2005 Jun;18(1):17-24.
- Huang H, Mackeen MM, Cook M, Oriero E, Locke E, Thézénas ML, Kessler BM, Nwakanma D, Casals-Pascual C. Proteomic identification of host and parasite biomarkers in saliva from patients with uncomplicated Plasmodium falciparum malaria. Malar J. 2012 May 28;11:178. doi: 10.1186/1475-2875-11-178.